



Article

# Melatonin MT<sub>1</sub> and MT<sub>2</sub> Receptors Exhibit Distinct Effects in the Modulation of Body Temperature across the Light/Dark Cycle

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**Abstract:** Melatonin (MLT) is a neurohormone that regulates many physiological functions including sleep, pain, thermoregulation, and circadian rhythms. MLT acts mainly through two G-protein-coupled receptors named MT<sub>1</sub> and MT<sub>2</sub>, but also through an MLT type-3 receptor (MT<sub>3</sub>). However, the role of MLT receptor subtypes in thermoregulation is still unknown. We have thus investigated the effects of selective and non-selective MLT receptor agonists/antagonists on body temperature (T<sub>b</sub>) in rats across the 12/12-h light–dark cycle. Rectal temperature was measured every 15 min from 4:00 a.m. to 9:30 a.m. and from 4:00 p.m. to 9:30 p.m., following subcutaneous injection of each compound at either 5:00 a.m. or 5:00 p.m. MLT (40 mg/kg) had no effect when injected at 5 a.m., whereas it decreased T<sub>b</sub> during the light phase only when injected at 5:00 p.m. This effect was blocked by the selective MT<sub>2</sub> receptor antagonist 4P-PDOT and the non-selective MT<sub>1</sub>/MT<sub>2</sub> receptor antagonist, luzindole, but not by the α<sub>1</sub>/MT<sub>3</sub> receptors antagonist prazosin. However, unlike MLT, neither the selective MT<sub>1</sub> receptor partial agonist UCM871 (14 mg/kg) nor the selective MT<sub>2</sub> partial agonist UCM924 (40 mg/kg) altered T<sub>b</sub> during the light phase. In contrast, UCM871 injected at 5:00 p.m. increased T<sub>b</sub> at the beginning of the dark phase, whereas UCM924 injected at 5:00 a.m. decreased T<sub>b</sub> at the end of the dark phase. These effects were blocked by luzindole and 4P-PDOT, respectively. The MT<sub>3</sub> receptor agonist GR135531 (10 mg/kg) did not affect T<sub>b</sub>. These data suggest that the simultaneous activation of both MT<sub>1</sub> and MT<sub>2</sub> receptors is necessary to regulate T<sub>b</sub> during the light phase, whereas in a complex but yet unknown manner, they regulate T<sub>b</sub> differently during the dark phase. Overall, MT<sub>1</sub> and MT<sub>2</sub> receptors display complementary but also distinct roles in modulating circadian fluctuations of T<sub>b</sub>.

**Keywords:** melatonin; MT<sub>1</sub> receptors; MT<sub>2</sub> receptors; MT<sub>3</sub> receptors; body temperature; light/dark cycle

## 1. Introduction

The maintenance of body temperature ( $T_b$ ) in mammals is critical for survival and internal homeostasis. The main brain structure involved in controlling  $T_b$  is the hypothalamus, which receives inputs from the thermoreceptors located in both the brain and the periphery. Depending on these inputs, homeostatic changes are subsequently induced, causing sweating or shivering [1]. In particular, the preoptic area (POA) and dorsomedial hypothalamus (DMH) are critical hypothalamic areas for thermoregulation. In these areas, based on the firing rate responses to changes in local brain temperature, electrophysiological recordings have shown three different neuronal populations: warm-sensitive neurons (~30%), cold-sensitive neurons (~6%), and insensitive neurons (~60%), [2,3]. Activation of the thermally-responsive GABAergic and glutamatergic neurons in the ventral part of the lateral preoptic nucleus (vLPO) and the dorsal part of the dorsomedial hypothalamus (DMD), respectively, decreases temperature, physical activity, and metabolic rate. On the contrary, GABAergic neurons in the DMD promote the increase of  $T_b$ , energy expenditure, and physical activity [4].

Recently, it has also been found that  $T_b$  can be internally regulated in a circadian manner by endogenous signals from other parts of the hypothalamus [4,5], specifically in the suprachiasmatic nucleus (SCN) [1,6,7]. The SCN regulates the circadian rhythmicity of several physiological responses [1], including the oscillatory decrease of the thermoregulatory threshold of heat production during day time and heat loss during night time in diurnal species [1,8,9]. However, the mechanisms underlying this circadian modulation of  $T_b$  remain unclear. Interestingly, the hypothalamus, including the SCN, DMH, and POA, are rich in melatonin (MLT)  $MT_1$  and  $MT_2$  receptors [7,10–13], and the activation of these MLT receptors modulates numerous physiological effects including the control of  $T_b$  [14].

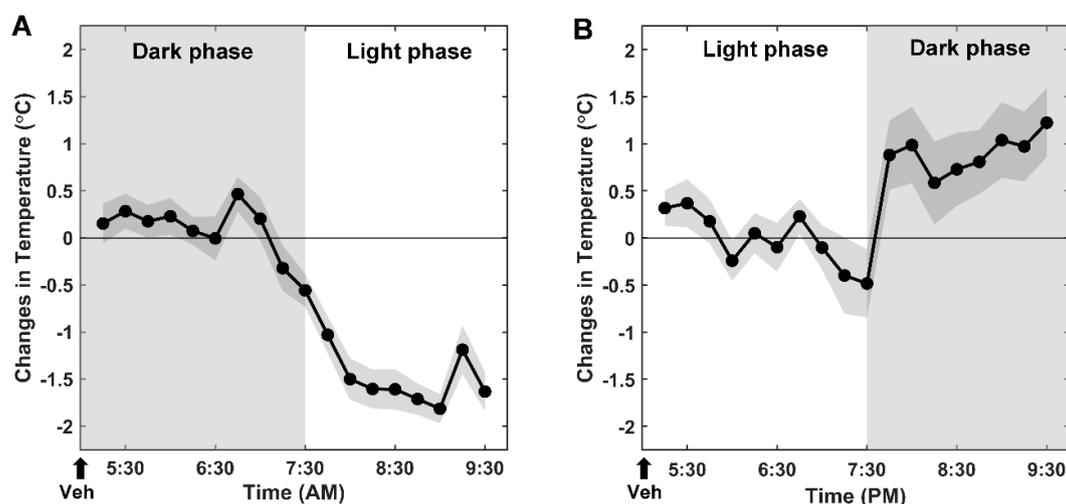
MLT is produced in the pineal gland, mostly during the dark phase in both diurnal and nocturnal species [12,15]. The circadian production and release of MLT are controlled by the SCN [16]. Most of the physiological effects of MLT result from the activation of two high-affinity G-protein coupled receptors (GPCRs),  $MT_1$  ( $pK_i = 10.09$ ) and  $MT_2$  ( $pK_i = 9.42$ ), both of which are widely expressed in the mammalian brain [7,10,17]. The specific localization of the two MLT receptor subtypes in different regions of the brain and/or neuronal populations [11,18–20] partially explains the selective and differential functional activity of the two MLT receptor subtypes, such as in sleep [18,21–23], anxiety [24], pain [25,26], circadian rhythms [27], and depression [28]. In addition to these high-affinity MLT receptors, another low-affinity MLT binding site, termed  $MT_3$  ( $pK_i = 6.0$ ), has been reported [29,30]. Given its both hydrophilic and lipophilic nature, MLT can easily pass through the cell membrane and bind nuclear receptors, including retinoic acid receptor-related orphan receptors (RORs) [31].

It is known that MLT decreases  $T_b$  during the night in diurnal species [1]. Clinical studies have shown that exogenous administration of MLT suppresses the physiological increase in  $T_b$  observed during daytime [32,33] and has hypothermic properties at the dose of 5 mg/kg [34–38]. Similar results have been demonstrated in preclinical studies in diurnal animals in which administration of MLT acted as a hypothermic agent in the active/light phase in fat sand rats and Marshall broiler chickens [39,40]. However, the neurobiological mechanism through which MLT exerts this hypothermic effect, as well as the selective contribution of the three MLT receptor subtypes, are yet to be investigated.

Therefore, we investigated modifications in  $T_b$  produced by selectively activating the three MLT receptor subtypes across the light–dark cycle in rats. To achieve this aim, we tested the effects of the selective  $MT_2$  receptor partial agonist N-[2-[(3-bromophenyl)-(4-fluorophenyl)amino]ethyl]acetamide (UCM924) ( $pK_{iMT_1} = 6.76$ ;  $pK_{iMT_2} = 9.27$ ) [41], the selective  $MT_1$  receptor partial agonist N-(2-[Methyl-[3-(4-phenylbutoxy)phenyl]amino]ethyl) acetamide (UCM871) ( $pK_{iMT_1} = 8.93$ ;  $pK_{iMT_2} = 7.04$ ) [42], and the  $MT_3$  receptor agonist 5-Methoxycarbonylamino-N-acetyltryptamine (GR135531) ( $pK_{iMT_3} = 29.5$ ) on  $T_b$ . The effects of UCM924, UCM871, and GR135531 were compared to those of MLT. In addition, selective and non-selective MLT receptor antagonists were also tested together with MLT and the other MLT receptor agonists/partial agonists to further dissect the MLT receptor subtypes involved in their thermoregulatory effects.

## 2. Results

In physiological conditions, as already known [1,9], there are changes in  $T_b$  between the light and the dark phase; in particular,  $T_b$  oscillations mostly occur during the phase shift (Figure 1). During the shift from the dark to the light phase, the  $T_b$  drops from an average of 38.45 to 37.5 °C after the light turns on (Figure 1A). The opposite occurs during the shift from the light to the dark phase when the light is turned off (Figure 1B).

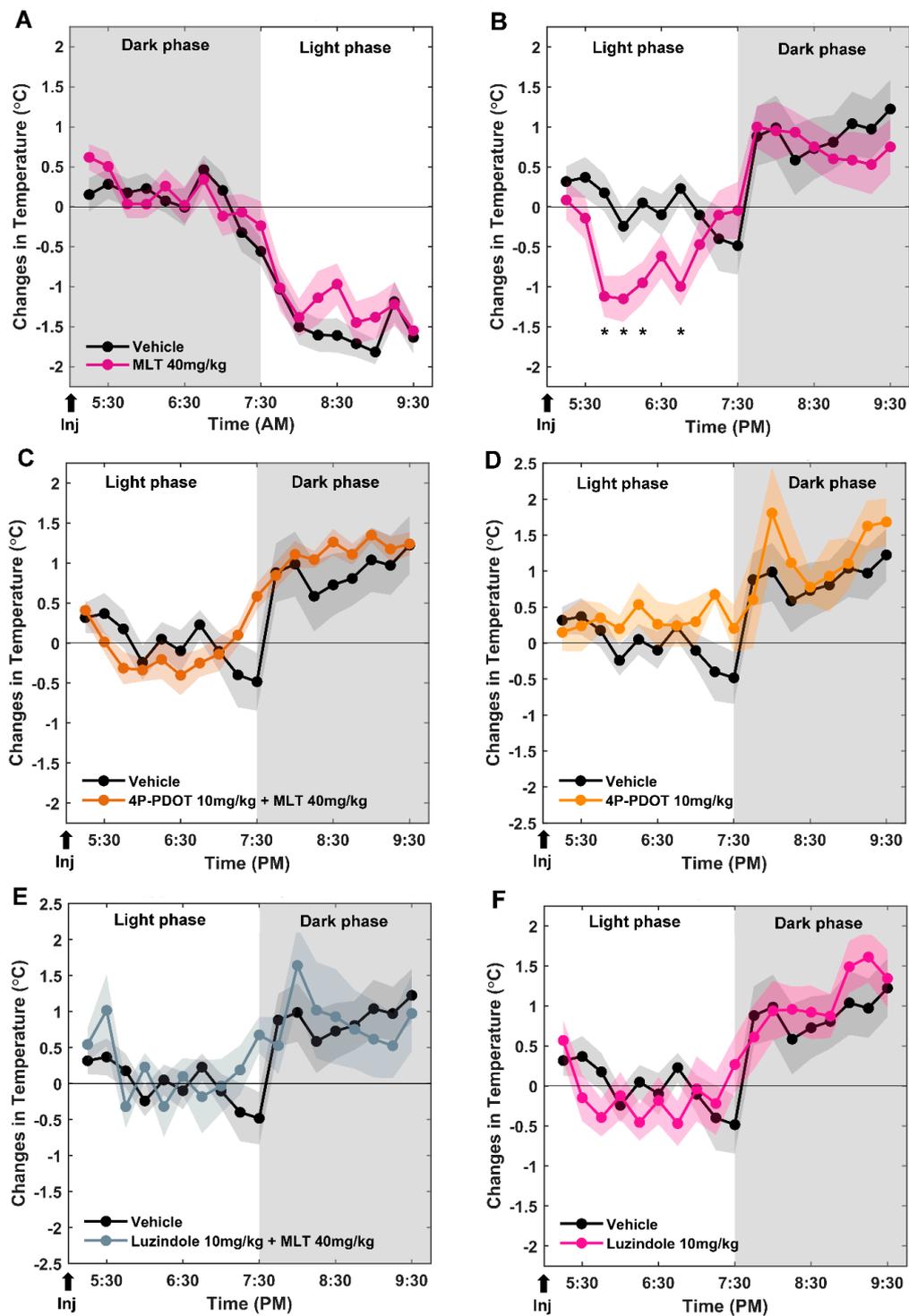


**Figure 1.** Body temperature ( $T_b$ ) changes during the light–dark cycle in rats exposed to a 12/12-h light–dark cycle. (A)  $T_b$  decreases during the transition from the dark phase to the light phase. (B)  $T_b$  increases during the transition from the light phase to the dark phase. Lights on at 7:30 a.m. and off at 7:30 p.m. Data represent mean value  $\pm$  SEM. Veh: s.c. injection of vehicle.

### 2.1. Effects of MLT Injected at the End of the Dark and of the Light Phases on $T_b$

As indicated in Figure 2A, the injection of MLT (40 mg/kg) at the end of the dark phase (5:00 a.m.) did not affect  $T_b$  during the end of the dark phase, the dark–light transition or the beginning of the light phase (two-way repeated measures ANOVA; interaction:  $F_{17,323} = 0.85$ ,  $p = 0.635$ ; treatment:  $F_{1,323} = 0.867$ ,  $p = 0.363$ ; time of the day:  $F_{17,323} = 31.835$ ,  $p < 0.001$ ).

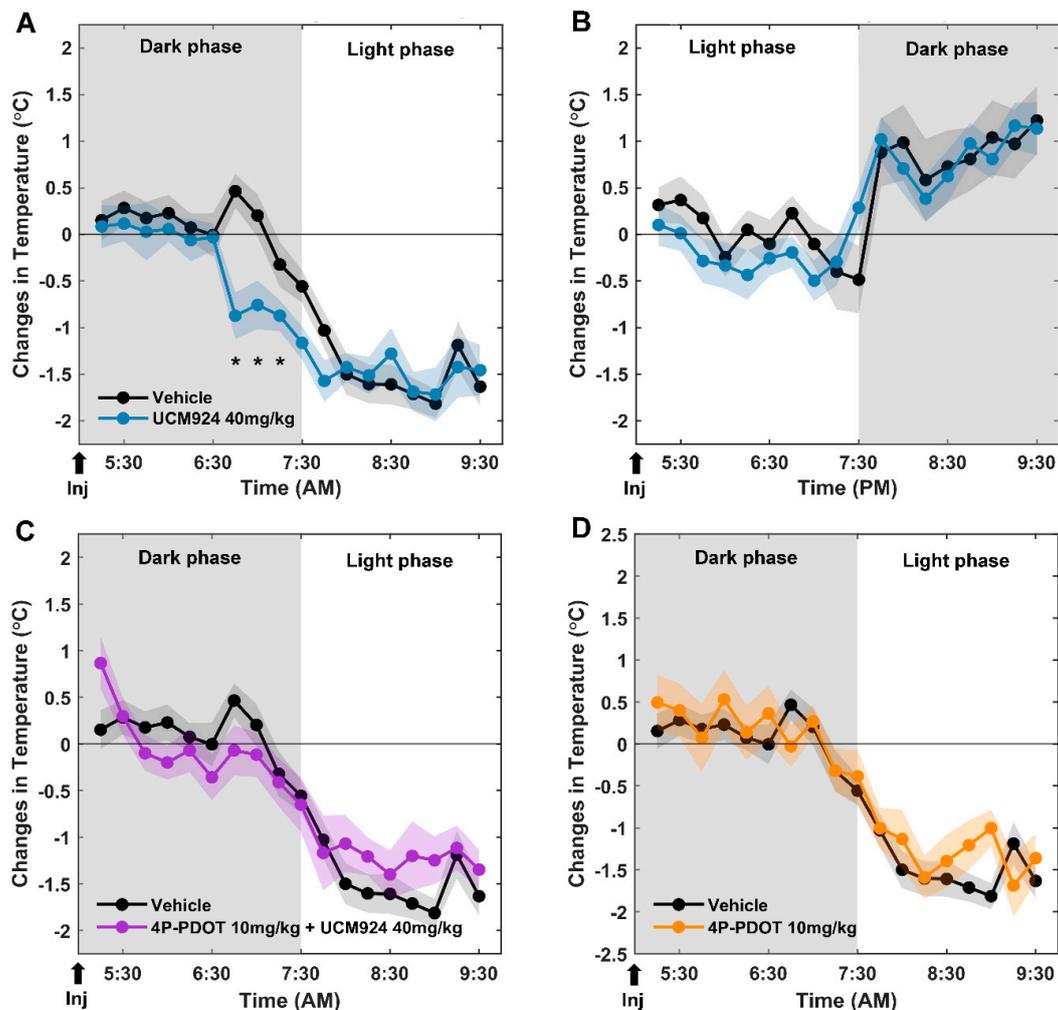
In contrast, when MLT (40 mg/kg) was injected at the end of the light phase (5:00 p.m.), it induced a significant decrease ( $p < 0.05$ ) in  $T_b$  from 5:45 p.m. to 6:45 p.m. that was close to the transition from the light to the dark phase (Figure 2B; interaction:  $F_{17,408} = 1.908$ ,  $p = 0.016$ ; treatment:  $F_{1,408} = 1.996$ ,  $p = 0.171$ ; time of the day:  $F_{17,408} = 10.658$ ,  $p < 0.001$ ). Importantly, we observed no further effects of MLT on  $T_b$  after the light–dark transition or during the beginning of the dark phase. The selective MT<sub>2</sub> receptor antagonist 4P-PDOT at a dose not affecting  $T_b$  (Figure 2D; interaction:  $F_{17,340} = 0.62$ ,  $p = 0.876$ ; treatment:  $F_{1,340} = 2.07$ ,  $p = 0.166$ ; time of the day:  $F_{17,340} = 4.86$ ,  $p < 0.001$ ) blocked the effects of MLT (Figure 2C; interaction:  $F_{17,391} = 1.448$ ,  $p = 0.111$ ; treatment:  $F_{1,391} = 0.22$ ,  $p = 0.643$ ; time of the day:  $F_{17,391} = 11.486$ ,  $p < 0.001$ ). Similarly, the pre-treatment with the selective MT<sub>1</sub>/MT<sub>2</sub> receptor antagonist luzindole at the dose not affecting  $T_b$  (Figure 2F; interaction:  $F_{17,408} = 1.144$ ,  $p = 0.309$ ; treatment:  $F_{1,408} = 0.012$ ,  $p = 0.912$ ; time of the day:  $F_{17,408} = 9.289$ ,  $p < 0.001$ ) also blocked the effects of MLT (Figure 2E; interaction:  $F_{17,289} = 0.989$ ,  $p = 0.47$ ; treatment:  $F_{1,289} = 0.11$ ,  $p = 0.745$ ; time of the day:  $F_{17,289} = 3.745$ ,  $p < 0.001$ ).



**Figure 2.** Changes in  $T_b$  after MLT administration (40 mg/kg) during the light and the dark phase. (A) MLT does not produce changes in  $T_b$  when administrated at 5:00 a.m. (B) MLT administrated during the dark phase (5:00 p.m.) decreases the  $T_b$  immediately after the administration compared with vehicle treated rats. (C) 4P-PDOT (10 mg/kg) pre-treatment blocks the effect of MLT on  $T_b$  during the light phase. (D) 4P-PDOT (10 mg/kg) injected during the light phase does not affect  $T_b$ . (E) Pre-treatment with luzindole (10 mg/kg) blocks the effect of MLT on  $T_b$  during the light phase. (F) luzindole (10 mg/kg) injected during the light phase does not affect  $T_b$ . Data are expressed as mean  $\pm$  SEM (graded shades). Lights on at 7:30 a.m. and off at 7:30 p.m. \*  $p < 0.05$  vs. vehicle; two-way ANOVA for repeated measures followed by Bonferroni *post hoc* test. Inj: s.c. injection of either vehicle, MLT, MLT + 4P-PDOT, 4P-PDOT, MLT + luzindole, or luzindole.

## 2.2. Effects of the Selective $MT_2$ Partial Agonist UCM924 Injected at the End of the Dark and of the Light Phases on $T_b$

As indicated in Figure 3A, the injection of UCM924 (40 mg/kg) at the end of the dark phase (5:00 a.m.) induced a significant decrease ( $p < 0.05$ ) in  $T_b$  immediately before the dark–light transition (from 6:45 a.m. to 7:30 a.m.), and did not affect  $T_b$  during the dark–light transition and at the beginning of the light phase (two-way repeated measures ANOVA; interaction:  $F_{17,289} = 0.2406$ ,  $p = 0.002$ ; treatment:  $F_{1,289} = 2.286$ ,  $p = 0.149$ ; time of the day:  $F_{17,289} = 30.597$ ,  $p < 0.001$ ).



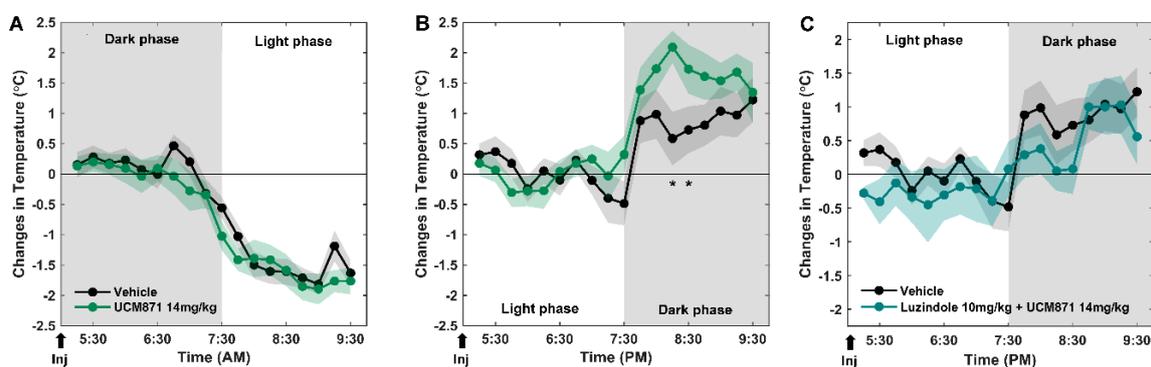
**Figure 3.** Changes in  $T_b$  after UCM924 (40 mg/kg, s.c.) treatment during the light and the dark phase. (A) UCM924 (40 mg/kg) administered at 5:00 a.m. (light phase) decreases  $T_b$  prior the light–dark shift compared with vehicle. (B) UCM924 (40 mg/kg) administered at 5:00 p.m. does not produce any change in the  $T_b$  compared with vehicle. (C) 4P-PDOT pre-treatment blocks the effect of UCM924 on  $T_b$  during the dark phase. (D) 4P-PDOT (10 mg/kg) injected during the dark phase does not affect  $T_b$ . Data are expressed as mean  $\pm$  SEM (graded shades). Lights on at 7:30 a.m. and off at 7:30 p.m. \*  $p < 0.05$  versus vehicle; two-way ANOVA for repeated measures followed by Bonferroni *post hoc* test. Inj: s.c. injection of either vehicle, UCM924, UCM924 + 4P-PDOT, or 4P-PDOT.

In contrast, when UCM924 (40 mg/kg) was injected during the light phase (5:00 p.m.), it did not affect  $T_b$  during the end of the light phase, the dark–light transition or the beginning of the dark phase (Figure 3B; two-way repeated measures ANOVA; interaction:  $F_{17,425} = 0.785$ ,  $p = 0.711$ ; treatment:  $F_{1,425} = 0.311$ ,  $p = 0.582$ ; time of the day:  $F_{17,425} = 9.618$ ,  $p < 0.001$ ).

The effects of UCM924 on  $T_b$  when injected during the dark phase were mediated by  $MT_2$  receptors since the pre-treatment with the selective  $MT_2$  receptor antagonist 4P-PDOT at a dose not affecting  $T_b$  (Figure 3D; interaction:  $F_{17,272} = 0.875$ ,  $p = 0.605$ ; treatment:  $F_{1,272} = 1.223$ ,  $p = 0.285$ ; time of the day:  $F_{17,272} = 22.848$ ,  $p < 0.001$ ) blocked the effects of UCM924 (Figure 3C; interaction:  $F_{17,272} = 1.922$ ,  $p = 0.016$ ; treatment:  $F_{1,272} = 0.035$ ,  $p = 0.821$ ; time of the day:  $F_{17,272} = 30.468$ ,  $p < 0.001$ ).

### 2.3. Effects of the Selective $MT_1$ Partial Agonist UCM871 Injected at the End of the Dark and of the Light Phases on $T_b$

As indicated in Figure 4A, the injection of UCM871 (14 mg/kg) at the end of the dark phase (5:00 a.m.) did not affect  $T_b$  during the end of dark phase, the dark–light transition or the beginning of light phase (two-way repeated measures ANOVA; interaction:  $F_{17,340} = 0.842$ ,  $p = 0.644$ ; treatment:  $F_{1,340} = 0.538$ ,  $p = 0.472$ ; time of the day:  $F_{17,340} = 44.622$ ,  $p < 0.001$ ).



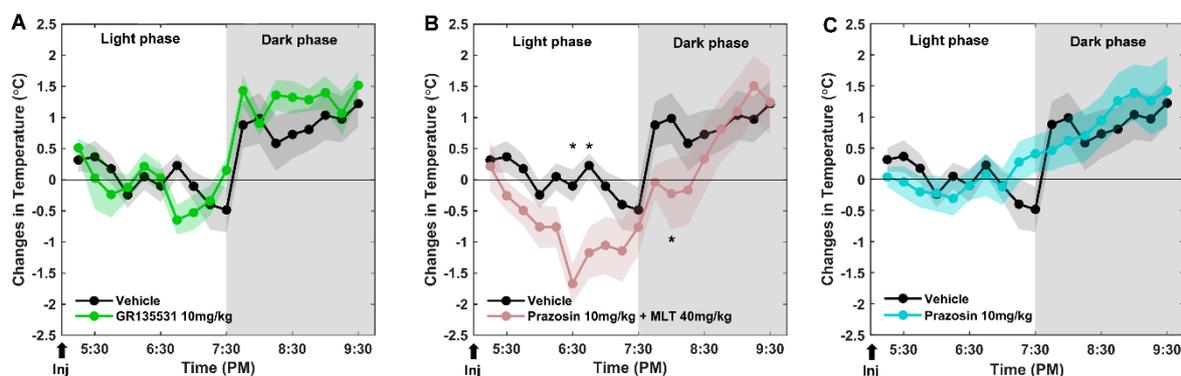
**Figure 4.** Changes in  $T_b$  after UCM871 (14 mg/kg, s.c.) treatment during the light and the dark phase. (A) UCM871 administered at 5:00 a.m. (dark phase) does not produce any change in the  $T_b$  compared with vehicle. (B) UCM871 administered at 5:00 p.m. (light phase) increases  $T_b$  after the light–dark transition compared with vehicle. (C) Luzindole pre-treatment blocks the effect of UCM871 on  $T_b$ . Data are expressed as mean  $\pm$  SEM (graded shades). Lights on at 7:30 a.m. and off at 7:30 p.m. \*  $p < 0.05$  versus vehicle; two-way ANOVA for repeated measures followed by Bonferroni *post hoc* test. Inj: s.c. injection of either vehicle, UCM871, or UCM871 + luzindole.

In contrast, when UCM871 (14 mg/kg) was injected during the light phase (5:00 p.m.), it induced a significant increase ( $p < 0.05$ ) in  $T_b$  from 8:15 p.m. to 8:45 p.m. (dark phase) (Figure 4B; interaction:  $F_{17,340} = 1.634$ ,  $p = 0.05$ ; treatment:  $F_{1,340} = 2.045$ ,  $p = 0.168$ ; time of the day:  $F_{17,340} = 10.42$ ,  $p < 0.001$ ). The effects of UCM871 on  $T_b$  when injected during the light phase were blocked by the pre-treatment with the  $MT_1/MT_2$  receptor antagonist luzindole (Figure 4C; interaction:  $F_{17,340} = 0.67$ ,  $p = 0.83$ ; treatment:  $F_{1,340} = 1.22$ ,  $p = 0.28$ ; time of the day:  $F_{17,340} = 4.88$ ,  $p < 0.001$ ) at a dose not affecting  $T_b$  (see Figure 2F; interaction:  $F_{17,408} = 1.144$ ,  $p = 0.309$ ; treatment:  $F_{1,408} = 0.012$ ,  $p = 0.912$ ; time of the day:  $F_{17,408} = 9.289$ ,  $p < 0.001$ ).

### 2.4. Effects of the Selective $MT_3$ Agonist GR135531 and Prazosin Injected at the End of the Light Phase on $T_b$

As indicated in Figure 5A, the injection of GR135531 (10 mg/kg) at the end of the light phase (5:00 p.m.) did not affect  $T_b$  during the end of the light phase, the dark–light transition or the beginning of dark phase (two-way repeated measures ANOVA; interaction:  $F_{17,306} = 0.94$ ,  $p = 0.527$ ; treatment:  $F_{1,306} = 0.342$ ,  $p = 0.566$ ; time of the day:  $F_{17,306} = 7.817$ ,  $p < 0.001$ ). The effects of MLT on  $T_b$ , when injected during the light phase, were not mediated by  $MT_3$  receptors, since the pre-treatment with the non-selective  $\alpha_1/MT_3$  antagonist prazosin at a dose not affecting  $T_b$  (Figure 5C, interaction:  $F_{17,357} = 1.01$ ,  $p = 0.446$ ; treatment:  $F_{1,357} = 0.022$ ,  $p = 0.882$ ; time of the day:  $F_{17,357} = 7.588$ ,  $p < 0.001$ ) did not block the effects of MLT (Figure 5B; interaction:  $F_{17,340} = 1.65$ ,  $p = 0.050$ ; treatment:  $F_{1,340} = 3.064$ ,  $p = 0.095$ ; time of the day:  $F_{17,340} = 10.086$ ,  $p < 0.001$ ). Interestingly, the treatment with prazosin plus

MLT induced a further decrease of  $T_b$  even during the dark phase at 8:00 p.m. (Figure 5B) that was not observed with MLT (Figure 2B) or prazosin (Figure 5C) alone.



**Figure 5.** (A) Changes in  $T_b$  after the administration of the MLT  $MT_3$  agonist GR135531 (10 mg/kg) during the dark phase. (B) Prazosin (10 mg/kg) pre-treatment does not block the effects of MLT on  $T_b$  during the light phase. (C) Prazosin (10 mg/kg) injected during the light phase does not affect  $T_b$ . Data are expressed as mean  $\pm$  SEM (graded shades). Lights off at 7:30 p.m. \*  $p < 0.05$  versus vehicle; two-way ANOVA for repeated measures followed by Bonferroni *post hoc* test. Inj: s.c. injection of either vehicle, GR135531, MLT + prazosin, or prazosin.

### 3. Discussion

In this study, we investigated the effects of MLT and its three receptors on  $T_b$  during the light and the dark phase for the first time. To achieve this aim, we used a pharmacological approach employing MLT, the selective  $MT_1$  receptor partial agonist UCM871, the selective  $MT_2$  receptor partial agonist UCM924, the  $MT_3$  receptor agonist GR135531, and selective/non-selective MLT receptor antagonists, including the  $MT_2$  selective antagonist 4P-PDOT, the  $MT_1/MT_2$  non-selective antagonist luzindole, and the  $MT_3/\alpha_1$  antagonist prazosin. The exogenous administration of MLT during the light phase decreased  $T_b$  immediately after the administration and before the light–dark phase shift, an effect blocked by both 4P-PDOT and luzindole. Interestingly, unlike MLT, neither UCM924 nor UCM871 produced a change in  $T_b$  during the light phase. In contrast, the selective  $MT_2$  partial agonist UCM924 administered at the end of the dark phase decreased  $T_b$  during the dark phase, just prior to the dark–light switch, whereas the selective  $MT_1$  partial agonist UCM871 injected at the end of the light phase increased  $T_b$  during the following dark phase. On the other hand,  $MT_3$  receptors did not seem to be involved in the regulation of  $T_b$ , since the  $MT_3$  receptor agonist GR135531 and the  $MT_3/\alpha_1$  antagonist prazosin alone had not produced any effect on  $T_b$ .

The rat circadian body temperature displays a cosine wave [39], showing a temperature that oscillates around the 35.6–36 °C during the light phase and around 37.8–38 °C during the dark phase [43]. The daily change in  $T_b$  shows a characteristic deviation at two different times, consistent with the switch between day (light phase) and night (dark phase) hours [44]. The present study replicates the same physiological  $T_b$  deviation that was previously reported in nocturnal rodents [43,44], with a daily  $T_b$  peak during the night, which is concomitant with the increase in the activity of the animals [45].

In nocturnal rodents,  $T_b$  peaks during night time when MLT levels are high, and decreases during the light phase when MLT levels are low. In contrast, in diurnal species,  $T_b$  regulation follows the reverse direction in relation to MLT levels, showing a  $T_b$  peak during the light phase [45] when circulating levels of MLT are very low ( $\sim 10$  pg/mL) [12,15,46]. However, the mechanism by which MLT regulates  $T_b$  has not been established.

The neuronal circuit controlling the regulation of  $T_b$  involves several structures including the SCN, SON, mPOA, and DMH [4,5]. Notably, the hypothalamus is an area rich in MLT receptors [11], and their expression may vary according to the phase and the time of the day [47–51].

Previous reports have shown that exogenous MLT administration during the light phase induced a decrease in  $T_b$  in both humans and rodents [32,35–37,39,52], although the active doses in humans are lower than those in rodents due to a significantly faster metabolism and very short half-life of MLT in the latter [53]. Our findings confirm that exogenous MLT influences  $T_b$ , and its effects are strictly dependent on the time of day: MLT reduces  $T_b$  only towards the end of the light phase and if administered during the light phase. In regard to its possible mechanism of action during the light phase, we found that both the selective  $MT_2$  antagonist 4P-PDOT and the non-selective  $MT_1/MT_2$  antagonist luzindole blocked the  $T_b$  reduction due to MLT, yet neither the selective  $MT_1$  partial agonist UCM871 nor the selective  $MT_2$  partial agonist UCM924 recapitulated the effects of MLT on  $T_b$ . These findings suggest that during the light phase, MLT needs to simultaneously activate both  $MT_1$  and  $MT_2$  receptors to modulate  $T_b$ . Indeed, the selective activation/inhibition of only  $MT_1$  or  $MT_2$  receptors did not affect  $T_b$  during the day. In contrast, UCM871 and UCM924 produced changes in  $T_b$  at different times of the dark phase and of opposite magnitude: UCM871 enhanced  $T_b$  just after the light–dark transition, whereas UCM924 decreased  $T_b$  just before the dark–light transition. Importantly, unlike UCM871 and UCM924, MLT did not induce any change in  $T_b$  during the dark phase. We previously observed a similar time-of-day-dependent effect of MLT on sleep [23]. However, it is interesting that when  $\alpha_1$  receptors/ $MT_3$  receptors were blocked by prazosin, MLT decreased  $T_b$  also during the dark phase. These complex findings observed during the dark phase are likely dependent on the fact that during the dark phase there is a significant increase in the endogenous levels of MLT, and thus the expression of the two MLT receptors [19], as well as the involvement of other receptors implicated in thermoregulation, such as  $\alpha_1$  adrenoceptors [54], probably vary.

Interestingly, it is now well recognized that MLT receptors can form  $MT_1/MT_2$  hetero-oligomers and also heteromers with other receptors, and from a functional point of view, their properties are different from those of the corresponding homomers [55,56]. Since MLT receptors as well as other receptors including  $\alpha_1$ -adrenoceptors are highly expressed in brain regions/nuclei involved in thermoregulation, we cannot exclude that some of the effects of MLT on  $T_b$  described here were mediated by these oligomers/heteromers. Future studies are needed to investigate the possible circadian variability in the formation and role of oligomers and/or heteromers of MLT receptors in hypothalamic nuclei regulating  $T_b$ . Similarly, the potential contribution of nuclear receptors, among which RORs that are also activated by MLT [31], is worth investigating.

The phase-dependent response of  $T_b$  to exogenous MLT may depend not only on the changes in the density of MLT receptors across the light–dark cycle, but also on the relative distribution and function of  $MT_1$  and  $MT_2$  receptors that control unique physiological responses in the brain, for example in sleep [21–23], anxiety [18,24], pain [25,26,57], and depression [28,58,59], and in the periphery, for example at the cardiovascular level [60,61].

In conclusion, we have investigated the role of MLT receptors in thermoregulation, and found that during the light phase  $T_b$  is affected only if both  $MT_1$  and  $MT_2$  receptors are simultaneously activated. Further, during the dark phase, a time-dependent effect was found in that the activation of  $MT_1$  and  $MT_2$  produces an increase and decrease of  $T_b$  respectively. No effects on  $T_b$  of the MLT  $MT_3$  receptor subtype were evidenced. However,  $MT_1$  and  $MT_2$  receptors control  $T_b$  in synergy with other receptors including  $\alpha_1$  adrenoceptors. These data further support the recent findings showing that the  $MT_1$  and  $MT_2$  receptors modulate physio-pathological functions in different and sometimes opposing ways, and in a time-of-day dependent manner. In particular,  $MT_1$  or  $MT_2$  agonists may be further tested for hypothermia or hyperthermia, respectively.

## 4. Materials and Methods

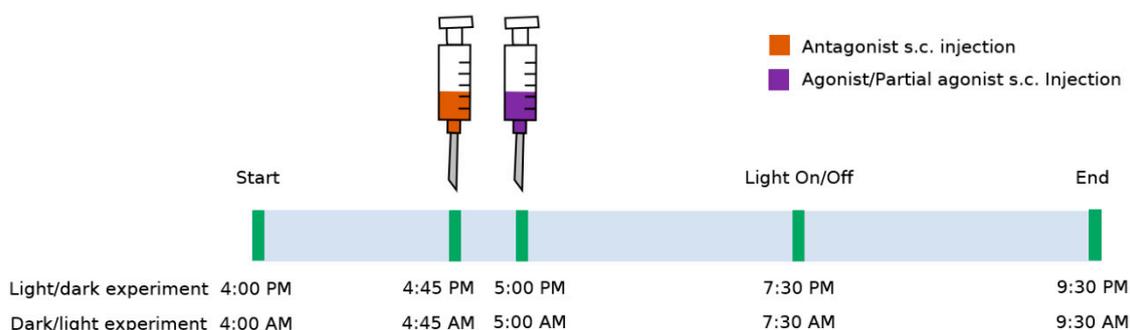
### 4.1. Animals

Male Wistar rats (200–250 g, Charles River) were used for behavioral tests. All animals were housed at constant room temperature (20 °C) and humidity under a 12/12-h light–dark cycle (lights

on at 7:30 a.m. and off at 7:30 p.m.) with food and water ad libitum. All experimental procedures were performed between 5:00 a.m. and 9:30 a.m. and between 5:00 p.m. to 9:30 p.m. The experimental protocol was approved by the Animal Ethics Committee (AUP#5253, McGill University, QC, Canada) and followed the ethical guidelines of the Canadian Institute of Health Research for animal care and scientific use.

#### 4.2. Drugs and Pharmacological Treatments

The N-(2-[Methyl-[3-(4-phenylbutoxy)phenyl]amino]ethyl)acetamide (UCM871, 14 mg/kg) [42] and N-(2-[3-bromophenyl)-(4-fluorophenyl)amino] ethyl)acetamide (UCM924, 40 mg/kg) [41] were synthesized by the University of Urbino, Italy, and by BioQuadrant Inc. (Montreal, Canada), respectively. Melatonin (40 mg/kg), luzindole (10 mg/kg), and prazosin hydrochloride (1-(4-Amino-6,7-dimethoxy-2-quinazoliny)-4-(2-furanylcarbonyl)piperazine hydrochloride; 10 mg/kg) were purchased from Sigma (St. Louis, MO, USA), and 4P-PDOT (4-phenyl-2-propionamidotetralin; 10 mg/kg) and GR135531 (5-Methoxycarbonylamino-N-acetyltryptamine, 10 mg/kg) from Tocris Bioscience (Ellisville, MO, USA). All drugs were dissolved in a vehicle composed of 70% dimethyl sulfoxide (MP Biochemicals, Solon, OH, USA) and 30% saline. The doses of UCM924, MLT, 4P-PDOT, and luzindole [23,25,26], as well as UCM871 [62], were chosen based on our previous experiments examining the potential pharmacological activity of these compounds. The doses of GR135531 and prazosin were based on the literature [63,64]. Drugs were injected subcutaneously (s.c.; 0.5 mL) 15 min before the beginning of the experiment: 5:00 a.m. for dark–light or 5:00 p.m. for light–dark testing. The selective MLT MT<sub>2</sub> receptor antagonist 4P-PDOT, the non-selective MLT MT<sub>1</sub>/MT<sub>2</sub> receptor antagonist luzindole, and the MT<sub>3</sub>/α<sub>1</sub> antagonist prazosin (pK<sub>i</sub> = 21.7) [65] were injected 15 min prior the agonist/partial agonist. Figure 6 describes the experimental protocol.



**Figure 6.** Schematic representation of the experimental design. Body temperature ( $T_b$ ) has been recorded every 15 min. For light–dark phase experiments, light was off at 7:30 p.m., whereas for dark–light phase experiments, light was on at 7:30 a.m. s.c.: subcutaneous.

#### 4.3. Assessment of Body Temperature

Body (rectal) temperature ( $T_b$ ) in awake animals was measured by goosing the animal using a Traceable Snap-in Module with probe (Fisher Science Education, S90862). The probe was inserted to a depth of 2 cm for no more than 10 s, whereas the tested individual was kept in a cotton bag. Animals were handled every day for five days before the experiments with the aim of habituating the animal to the testing procedure and thus minimizing the associated stress.

#### 4.4. Statistical Analysis

Data analysis was conducted using the SigmaPlot statistical software version 13 (Systat Software, Inc.). After controlling for the normal distribution of the data, a two-way ANOVA for repeated measures was used to analyze the data using treatments (between) and testing time (within) as factors. Post hoc analyses were performed using the Bonferroni test for multiple comparisons. The effect of vehicle was compared with that of the different agonists/partial agonists, the antagonists alone, or the

agonist/partial agonist plus the antagonist. All data were expressed as mean  $\pm$  SEM.  $p < 0.05$  was considered significant. All figures were made using MATLAB software.

Temperature values were normalized as follow: [Temperature at time X – Average of Temperature (4:00 a.m./p.m. to 5:00 a.m./p.m.)]/[Average of Temperature (4:00 a.m./p.m. to 5:00 a.m./p.m.)]. “Time X” indicates any time after the injection.

**Author Contributions:** Conceptualization, G.G.; Data curation, M.L.-C., S.H.M., and S.C.; Formal analysis, M.L.-C. and S.C.; Investigation, M.L.-C., S.H.M., L.P., and D.D.G.; Project administration, G.G.; Resources, A.B. and G.S.; Supervision, G.G. and S.C.; Writing—original draft, M.L.-C. and S.H.M.; Writing—review & editing, G.G. and S.C.

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## References

1. Krauchi, K.; Deboer, T. The interrelationship between sleep regulation and thermoregulation. *Front. Biosci.* **2010**, *15*, 604–625. [[CrossRef](#)]
2. Boulant, J.A. Role of the preoptic-anterior hypothalamus in thermoregulation and fever. *Clin. Infect. Dis.* **2000**, *31*, S157–S161. [[CrossRef](#)] [[PubMed](#)]
3. Boulant, J.A. Counterpoint: Heat-induced membrane depolarization of hypothalamic neurons: An unlikely mechanism of central thermosensitivity. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **2006**, *290*, R1481–R1484. [[PubMed](#)]
4. Zhao, Z.D.; Yang, W.Z.; Gao, C.; Fu, X.; Zhang, W.; Zhou, Q.; Chen, W.; Ni, X.; Lin, J.K.; Yang, J.; et al. A hypothalamic circuit that controls body temperature. *Proc. Natl. Acad. Sci. USA* **2017**, *114*, 2042–2047. [[CrossRef](#)] [[PubMed](#)]
5. Liedtke, W.B. Deconstructing mammalian thermoregulation. *Proc. Natl. Acad. Sci. USA* **2017**, *114*, 1765–1767. [[CrossRef](#)]
6. Krauchi, K.; Cajochen, C.; Pache, M.; Flammer, J.; Wirz-Justice, A. Thermoregulatory effects of melatonin in relation to sleepiness. *Chronobiol. Int.* **2006**, *23*, 475–484. [[CrossRef](#)] [[PubMed](#)]
7. Ng, K.Y.; Leong, M.K.; Liang, H.; Paxinos, G. Melatonin receptors: Distribution in mammalian brain and their respective putative functions. *Brain Struct. Funct.* **2017**, *222*, 2921–2939. [[CrossRef](#)]
8. Benstaali, C.; Mailloux, A.; Bogdan, A.; Auzéby, A.; Touitou, Y. Circadian rhythms of body temperature and motor activity in rodents their relationships with the light-dark cycle. *Life Sci.* **2001**, *68*, 2645–2656. [[CrossRef](#)]
9. Krauchi, K.; Wirz-Justice, A. Circadian rhythm of heat production, heart rate, and skin and core temperature under unmasking conditions in men. *Am. J. Physiol.* **1994**, *267*, R819–R829. [[CrossRef](#)] [[PubMed](#)]
10. Dubocovich, M.L.; Markowska, M. Functional MT1 and MT2 melatonin receptors in mammals. *Endocrine* **2005**, *27*, 101–110. [[CrossRef](#)]
11. Lacoste, B.; Angeloni, D.; Dominguez-Lopez, S.; Calderoni, S.; Mauro, A.; Fraschini, F.; Descarries, L.; Gobbi, G. Anatomical and cellular localization of melatonin MT1 and MT2 receptors in the adult rat brain. *J. Pineal. Res.* **2015**, *58*, 397–417. [[CrossRef](#)] [[PubMed](#)]
12. Pandi-Perumal, S.R.; Srinivasan, V.; Maestroni, G.J.; Cardinali, D.P.; Poeggeler, B.; Hardeland, R. Melatonin: Nature's most versatile biological signal? *FEBS J.* **2006**, *273*, 2813–2838. [[CrossRef](#)] [[PubMed](#)]
13. Wu, Y.H.; Zhou, J.N.; Balesar, R.; Unmehopa, U.; Bao, A.; Jockers, R.; Van Heerikhuizen, J.; Swaab, D.F. Distribution of MT1 melatonin receptor immunoreactivity in the human hypothalamus and pituitary gland: Colocalization of MT1 with vasopressin, oxytocin, and corticotropin-releasing hormone. *J. Comp. Neurol.* **2006**, *499*, 897–910. [[CrossRef](#)] [[PubMed](#)]
14. Claustrat, B.; Leston, J. Melatonin: Physiological effects in humans. *Neurochirurgie* **2015**, *61*, 77–84. [[CrossRef](#)] [[PubMed](#)]
15. Arendt, J. Melatonin. *Clin. Endocrinol.* **1988**, *29*, 205–229. [[CrossRef](#)]

16. Klein, D.C.; Moore, R.Y.; Reppert, S.M. *Suprachiasmatic Nucleus: The Mind's Clock*; Oxford University Press: Oxford, UK, 1991.
17. Hardeland, R.; Poeggeler, B. Melatonin and synthetic melatonergic agonists: Actions and metabolism in the central nervous system. *Cent. Nerv. Syst. Agents Med. Chem.* **2012**, *12*, 189–216. [[CrossRef](#)] [[PubMed](#)]
18. Comai, S.; Gobbi, G. Unveiling the role of melatonin MT2 receptors in sleep, anxiety and other neuropsychiatric diseases: A novel target in psychopharmacology. *J. Psychiatry Neurosci.* **2014**, *39*, 6–21. [[CrossRef](#)] [[PubMed](#)]
19. Gobbi, G.; Comai, S. Differential Function of Melatonin MT1 and MT2 Receptors in REM and NREM Sleep. *Front. Endocrinol (Lausanne)* **2019**, *10*, 87. [[CrossRef](#)]
20. Gobbi, G.; Comai, S. Sleep well. Untangling the role of melatonin MT1 and MT2 receptors in sleep. *J. Pineal Res.* **2019**, *66*, e12544. [[CrossRef](#)]
21. Comai, S.; Ochoa-Sanchez, R.; Gobbi, G. Sleep-wake characterization of double MT(1)/MT(2) receptor knockout mice and comparison with MT(1) and MT(2) receptor knockout mice. *Behav. Brain Res.* **2013**, *243*, 231–238. [[CrossRef](#)]
22. Ochoa-Sanchez, R.; Comai, S.; Lacoste, B.; Bambico, F.R.; Dominguez-Lopez, S.; Spadoni, G.; Rivara, S.; Bedini, A.; Angeloni, D.; Fraschini, F.; et al. Promotion of non-rapid eye movement sleep and activation of reticular thalamic neurons by a novel MT2 melatonin receptor ligand. *J. Neurosci.* **2011**, *31*, 18439–18452. [[CrossRef](#)]
23. Ochoa-Sanchez, R.; Comai, S.; Spadoni, G.; Bedini, A.; Tarzia, G.; Gobbi, G. Melatonin, selective and non-selective MT1/MT2 receptors agonists: Differential effects on the 24-h vigilance states. *Neurosci. Lett.* **2014**, *561*, 156–161. [[CrossRef](#)]
24. Ochoa-Sanchez, R.; Rainer, Q.; Comai, S.; Spadoni, G.; Bedini, A.; Rivara, S.; Fraschini, F.; Mor, M.; Tarzia, G.; Gobbi, G. Anxiolytic effects of the melatonin MT(2) receptor partial agonist UCM765: Comparison with melatonin and diazepam. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **2012**, *39*, 318–325. [[CrossRef](#)]
25. Lopez-Canul, M.; Comai, S.; Dominguez-Lopez, S.; Granados-Soto, V.; Gobbi, G. Antinociceptive properties of selective MT(2) melatonin receptor partial agonists. *Eur. J. Pharmacol* **2015**, *764*, 424–432. [[CrossRef](#)] [[PubMed](#)]
26. Lopez-Canul, M.; Palazzo, E.; Dominguez-Lopez, S.; Luongo, L.; Lacoste, B.; Comai, S.; Angeloni, D.; Fraschini, F.; Boccella, S.; Spadoni, G.; et al. Selective melatonin MT2 receptor ligands relieve neuropathic pain through modulation of brainstem descending antinociceptive pathways. *Pain* **2015**, *156*, 305–317. [[CrossRef](#)]
27. Pevet, P. Melatonin receptors as therapeutic targets in the suprachiasmatic nucleus. *Expert Opin. Ther. Targets* **2016**, *20*, 1209–1218. [[CrossRef](#)] [[PubMed](#)]
28. Comai, S.; Ochoa-Sanchez, R.; Dominguez-Lopez, S.; Bambico, F.R.; Gobbi, G. Melancholic-Like behaviors and circadian neurobiological abnormalities in melatonin MT1 receptor knockout mice. *Int. J. Neuropsychopharmacol.* **2015**, *18*, pyu075. [[CrossRef](#)]
29. Nosjean, O.; Ferro, M.; Coge, F.; Beauverger, P.; Henlin, J.M.; Lefoulon, F.; Fauchere, J.L.; Delagrangé, P.; Canet, E.; Boutin, J.A. Identification of the melatonin-binding site MT3 as the quinone reductase 2. *J. Biol Chem* **2000**, *275*, 31311–31317. [[CrossRef](#)]
30. Slominski, R.M.; Reiter, R.J.; Schlabritz-Loutsevitch, N.; Ostrom, R.S.; Slominski, A.T. Melatonin membrane receptors in peripheral tissues: Distribution and functions. *Mol. Cell Endocrinol.* **2012**, *351*, 152–166. [[CrossRef](#)] [[PubMed](#)]
31. Hardeland, R. Melatonin: Signaling mechanisms of a pleiotropic agent. *Biofactors* **2009**, *35*, 183–192. [[CrossRef](#)]
32. Cagnacci, A.; Elliott, J.A.; Yen, S.S. Melatonin: A major regulator of the circadian rhythm of core temperature in humans. *J. Clin. Endocrinol Metab* **1992**, *75*, 447–452.
33. Hughes, R.J.; Badia, P. Sleep-promoting and hypothermic effects of daytime melatonin administration in humans. *Sleep* **1997**, *20*, 124–131. [[CrossRef](#)]
34. Arendt, J.; Skene, D.J. Melatonin as a chronobiotic. *Sleep Med. Rev.* **2005**, *9*, 25–39. [[CrossRef](#)]
35. Cagnacci, A. Melatonin in relation to physiology in adult humans. *J. Pineal Res.* **1996**, *21*, 200–213. [[CrossRef](#)]
36. Cagnacci, A.; Soldani, R.; Romagnolo, C.; Yen, S.S. Melatonin-induced decrease of body temperature in women: A threshold event. *Neuroendocrinology* **1994**, *60*, 549–552. [[CrossRef](#)]
37. Gilbert, S.S.; van den Heuvel, C.J.; Dawson, D. Daytime melatonin and temazepam in young adult humans: Equivalent effects on sleep latency and body temperatures. *J. Physiol.* **1999**, *514*, 905–914. [[CrossRef](#)]

38. Marrin, K.; Drust, B.; Gregson, W.; Atkinson, G. A meta-analytic approach to quantify the dose-response relationship between melatonin and core temperature. *Eur. J. Appl. Physiol.* **2013**, *113*, 2323–2329. [[CrossRef](#)]
39. Schwimmer, H.; Mursu, N.; Haim, A. Effects of light and melatonin treatment on body temperature and melatonin secretion daily rhythms in a diurnal rodent, the fat sand rat. *Chronobiol. Int.* **2010**, *27*, 1401–1419. [[CrossRef](#)]
40. Sinkalu, V.O.; Ayo, J.O.; Adelaiye, A.B.; Hambolu, J.O. Ameliorative effects of melatonin administration and photoperiods on diurnal fluctuations in cloacal temperature of Marshall broiler chickens during the hot dry season. *Int. J. Biometeorol.* **2015**, *59*, 79–87. [[CrossRef](#)]
41. Rivara, S.; Vacondio, F.; Fioni, A.; Silva, C.; Carmi, C.; Mor, M.; Lucini, V.; Pannacci, M.; Caronno, A.; Scaglione, F.; et al. N-(Anilinoethyl)amides: Design and synthesis of metabolically stable, selective melatonin receptor ligands. *ChemMedChem* **2009**, *4*, 1746–1755. [[CrossRef](#)]
42. Rivara, S.; Pala, D.; Lodola, A.; Mor, M.; Lucini, V.; Dugnani, S.; Scaglione, F.; Bedini, A.; Lucarini, S.; Tarzia, G.; et al. MT1-selective melatonin receptor ligands: Synthesis, pharmacological evaluation, and molecular dynamics investigation of N-[(3-O-substituted)anilino]alkylamides. *ChemMedChem* **2012**, *7*, 1954–1964. [[CrossRef](#)]
43. De Vries, J.; Strubbe, J.H.; Wildering, W.C.; Gorter, J.A.; Prins, A.J. Patterns of body temperature during feeding in rats under varying ambient temperatures. *Physiol. Behav.* **1993**, *53*, 229–235. [[CrossRef](#)]
44. Brieese, E. Normal body temperature of rats: The setpoint controversy. *Neurosci. Biobehav. Rev.* **1998**, *22*, 427–436. [[CrossRef](#)]
45. McElhinny, T.L.; Smale, L.; Holekamp, K.E. Patterns of body temperature, activity, and reproductive behavior in a tropical murid rodent, *Arvicanthis niloticus*. *Physiol. Behav.* **1997**, *62*, 91–96. [[CrossRef](#)]
46. Karasek, M. Melatonin, human aging, and age-related diseases. *Exp. Gerontol.* **2004**, *39*, 1723–1729. [[CrossRef](#)]
47. Pinato, L.; Ramos, D.; Hataka, A.; Rossignoli, P.S.; Granado, M.D.J.; Mazzetto, M.C.; Campos, L.M.G. Day/night expression of MT1 and MT2 receptors in hypothalamic nuclei of the primate *Sapajus apella*. *J. Chem. Neuroanat.* **2017**, *81*, 10–17. [[CrossRef](#)]
48. Waly, N.E.; Hallworth, R. Circadian Pattern of Melatonin MT1 and MT2 Receptor Localization in the Rat Suprachiasmatic Nucleus. *J. Circad. Rhythms* **2015**, *13*, 1. [[CrossRef](#)]
49. Masana, M.I.; Benloucif, S.; Dubocovich, M.L. Circadian rhythm of mt1 melatonin receptor expression in the suprachiasmatic nucleus of the C3H/HeN mouse. *J. Pineal Res.* **2000**, *28*, 185–192. [[CrossRef](#)]
50. Odo, M.; Koh, K.; Takada, T.; Yamashita, A.; Narita, M.; Kuzumaki, N.; Ikegami, D.; Sakai, H.; Iseki, M.; Inada, E.; et al. Changes in circadian rhythm for mRNA expression of melatonin 1A and 1B receptors in the hypothalamus under a neuropathic pain-like state. *Synapse* **2014**, *68*, 153–158. [[CrossRef](#)]
51. Poirel, V.J.; Masson-Pevet, M.; Pevet, P.; Gauer, F. MT1 melatonin receptor mRNA expression exhibits a circadian variation in the rat suprachiasmatic nuclei. *Brain Res.* **2002**, *946*, 64–71. [[CrossRef](#)]
52. Cagnacci, A.; Krauchi, K.; Wirz-Justice, A.; Volpe, A. Homeostatic versus circadian effects of melatonin on core body temperature in humans. *J. Biol. Rhythms* **1997**, *12*, 509–517. [[CrossRef](#)]
53. Yeleswaram, K.; McLaughlin, L.G.; Knipe, J.O.; Schabdach, D. Pharmacokinetics and oral bioavailability of exogenous melatonin in preclinical animal models and clinical implications. *J. Pineal Res.* **1997**, *22*, 45–51. [[CrossRef](#)]
54. Alam, M.N.; Mallick, B.N. Role of lateral preoptic area alpha-1 and alpha-2 adrenoceptors in sleep-wakefulness and body temperature regulation. *Brain Res. Bull.* **1994**, *35*, 171–177. [[CrossRef](#)]
55. Oishi, A.; Cecon, E.; Jockers, R. Melatonin Receptor Signaling: Impact of Receptor Oligomerization on Receptor Function. *Int. Rev. Cell Mol. Biol.* **2018**, *338*, 59–77. [[PubMed](#)]
56. Kamal, M.; Gbahou, F.; Guillaume, J.L.; Daulat, A.M.; Benleulmi-Chaachoua, A.; Luka, M.; Chen, P.; Kalbasi Anaraki, D.; Baroncini, M.; Mannoury la Cour, C.; et al. Convergence of melatonin and serotonin (5-HT) signaling at MT2/5-HT2C receptor heteromers. *J. Biol. Chem.* **2015**, *290*, 11537–11546. [[CrossRef](#)]
57. Posa, L.; De Gregorio, D.; Gobbi, G.; Comai, S. Targeting Melatonin MT2 Receptors: A Novel Pharmacological Avenue for Inflammatory and Neuropathic Pain. *Curr. Med. Chem.* **2018**, *25*, 3866–3882. [[CrossRef](#)] [[PubMed](#)]
58. Liu, J.; Clough, S.J.; Dubocovich, M.L. Role of the MT1 and MT2 melatonin receptors in mediating depressive- and anxiety-like behaviors in C3H/HeN mice. *Genes Brain Behav.* **2017**, *16*, 546–553. [[CrossRef](#)]
59. Comai, S.; Lopez-Canul, M.; De Gregorio, D.; Posner, A.; Ettaoussi, M.; Guarnieri, F.; Gobbi, G. Melatonin MT1 receptor as a novel target in neuropsychopharmacology: MT1 ligands, pathophysiological and therapeutic implications and perspectives. *Pharmacol. Res.* **2019**. [[CrossRef](#)]

60. Doolen, S.; Krause, D.N.; Dubocovich, M.L.; Duckles, S.P. Melatonin mediates two distinct responses in vascular smooth muscle. *Eur. J. Pharmacol.* **1998**, *345*, 67–69. [[CrossRef](#)]
61. Pandi-Perumal, S.R.; BaHammam, A.S.; Ojike, N.I.; Akinseye, O.A.; Kendzerska, T.; Buttoo, K.; Dhandapany, P.S.; Brown, G.M.; Cardinali, D.P. Melatonin and Human Cardiovascular Disease. *J. Cardiovasc. Pharmacol. Ther.* **2017**, *22*, 122–132. [[CrossRef](#)] [[PubMed](#)]
62. Comai, S.; Posa, L.; Ochoa-Sanchez, R.; Spadoni, S.; Gobbi, G. Neuropsychopharmacological properties of novel melatonin MT1 receptor ligands. *Eur. Neuropsychopharmacol.* **2017**, *27*. [[CrossRef](#)]
63. Pintor, J.; Pelaez, T.; Hoyle, C.H.; Peral, A. Ocular hypotensive effects of melatonin receptor agonists in the rabbit: Further evidence for an MT3 receptor. *Br. J. Pharmacol.* **2003**, *138*, 831–836. [[CrossRef](#)] [[PubMed](#)]
64. Zurowski, D.; Nowak, L.; Machowska, A.; Wordliczek, J.; Thor, P.J. Exogenous melatonin abolishes mechanical allodynia but not thermal hyperalgesia in neuropathic pain. The role of the opioid system and benzodiazepine-gabaergic mechanism. *J. Physiol. Pharmacol.* **2012**, *63*, 641–647.
65. Molinari, E.J.; North, P.C.; Dubocovich, M.L. 2-[125I]iodo-5-methoxycarbonylamino-N-acetyltryptamine: A selective radioligand for the characterization of melatonin ML2 binding sites. *Eur. J. Pharmacol.* **1996**, *301*, 159–168. [[CrossRef](#)]



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